

AB7500Fast USERS MANUAL

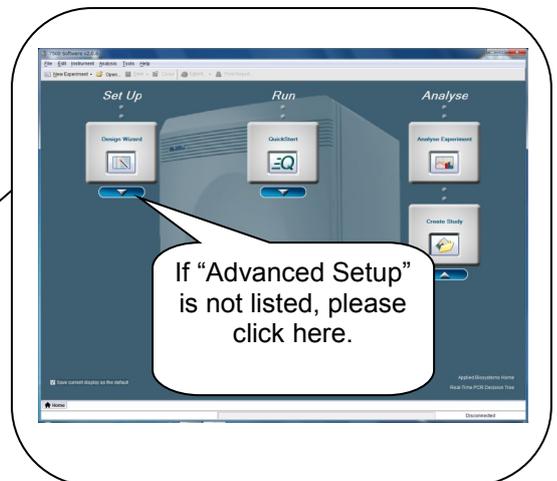
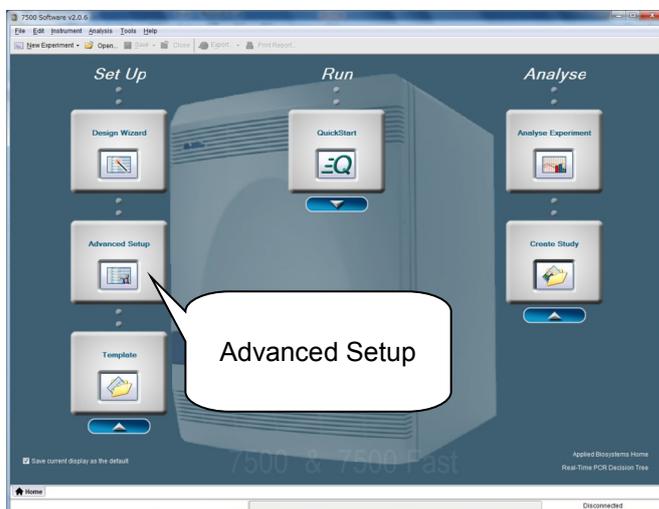
NOTES

- This system can use a 100 ul-reaction plate (tube). Don't use a 200 ul-plate (tube). That will cause a fatal error of this Real-Time PCR system.
- Don't perform data analysis and data transfer during running an experiment. There is a risk of stopping data collection.
- Don't write anything on the reaction plate (tube) with felt-tip pen. That will cause the dirt of the heat block and the heat cover.
- You should be responsible for your data safety. Some accidents cause the loss of the data saved to the hard drive.
- Make sure that the temperature of the heat block begin to rise. If the temperature isn't rising, please operate in the following procedure.
 - 1) Stop running
 - 2) Close the software and delete the file.
 - 3) Restart the PC and the device(7500Fast)
 - 4) Create new file
 - 5) Start RUN

1) Starting the 7500 Fast SDS software



2) Select "Advanced Setup"



3) Set up the Experimental Properties.

Experiment: 20151214name Type: Standard Curve Reagents: SYBR® Green Reagents START RUN

Experiment Properties

How do you want to identify the experiment?

Experiment Name: 20151214name

Barcode (Optional):

User Name (Optional):

Comments (Optional):

Which instrument are you using to run the experiment?

7500 (96 Wells) 7500 Fast (96 Wells)

Set up, run, and analyze an experiment using a fast cycling instrument.

Which type of experiment do you want to set up?

Quantitation - Standard Curve Quantitation - Relative Standard Curve Quantitation - Comparative Ct

Melt Curve Genotyping Presence/Absence

Standards to determine the absolute quantity of target nucleic acid sequence in samples.

Which reagents do you want to use to detect the target sequence?

TaqMan® Reagents SYBR® Green Reagents Other

The PCR reactions contain primers designed to amplify the target sequence and SYBR® Green dye to detect double-stranded DNA.

Include Melt Curve

Which amplicon do you want to use in the instrument run?

Standard (~ 2 hours to complete a run) Fast (~ 40 minutes to complete a run)

When you select "SYBR Green Reagent", this check box (include Melt Curve) is displayed.

Date + free word
Ex) 20151224ocu

You must select the "7500 Fast (96 wells)".

Select the fluorescent reagent

Please check the package insert of the reagent you used.

When you don't have enough time to define plate setup, you can start run after setting this "Experimental Properties".

4) Define Plate Setup

4)-1 Define Target and Samples

Experiment: 20151214name etc Type: Standard Curve Reagents: TaqMan® Reagents START RUN

Define Targets and Samples Assign Targets and Samples

Define Targets

Add New Target Add Saved Target Save Target Delete Target

Target Name	Reporter	Quencher	Colour
Target 1	FAM	NFQ-MGB	
Target 2	FAM	NFQ-MGB	

Define Samples

Add New Sample Add Saved Sample Save Sample Delete Sample

Sample Name	Color
Sample 1	
Sample 2	
Sample 3	
Sample 4	
Sample 5	
Sample 6	

Define Biological Groups

Instructions: For each biological replicate group in the reaction plate, click Add Biological Group, then click Assign Targets and Samples.

Add Biological Group Delete Biological Group

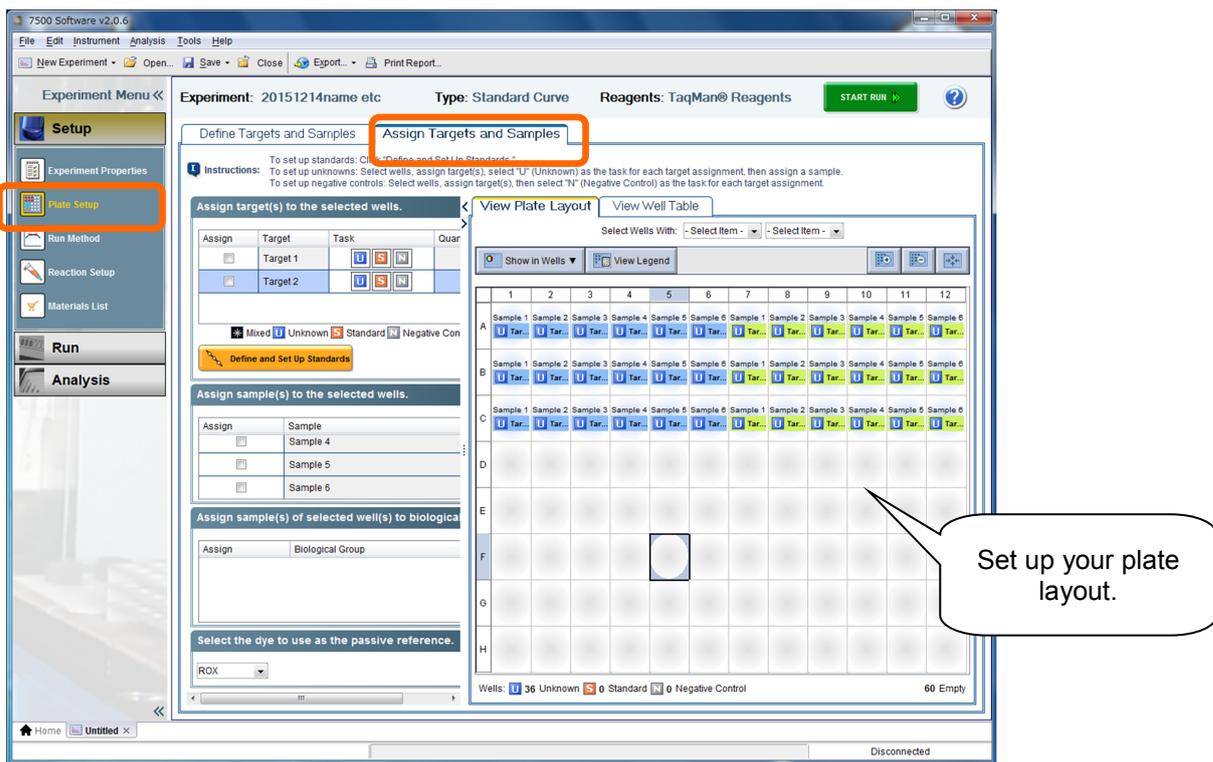
Biological Group Name	Color

Assign Targets and Samples

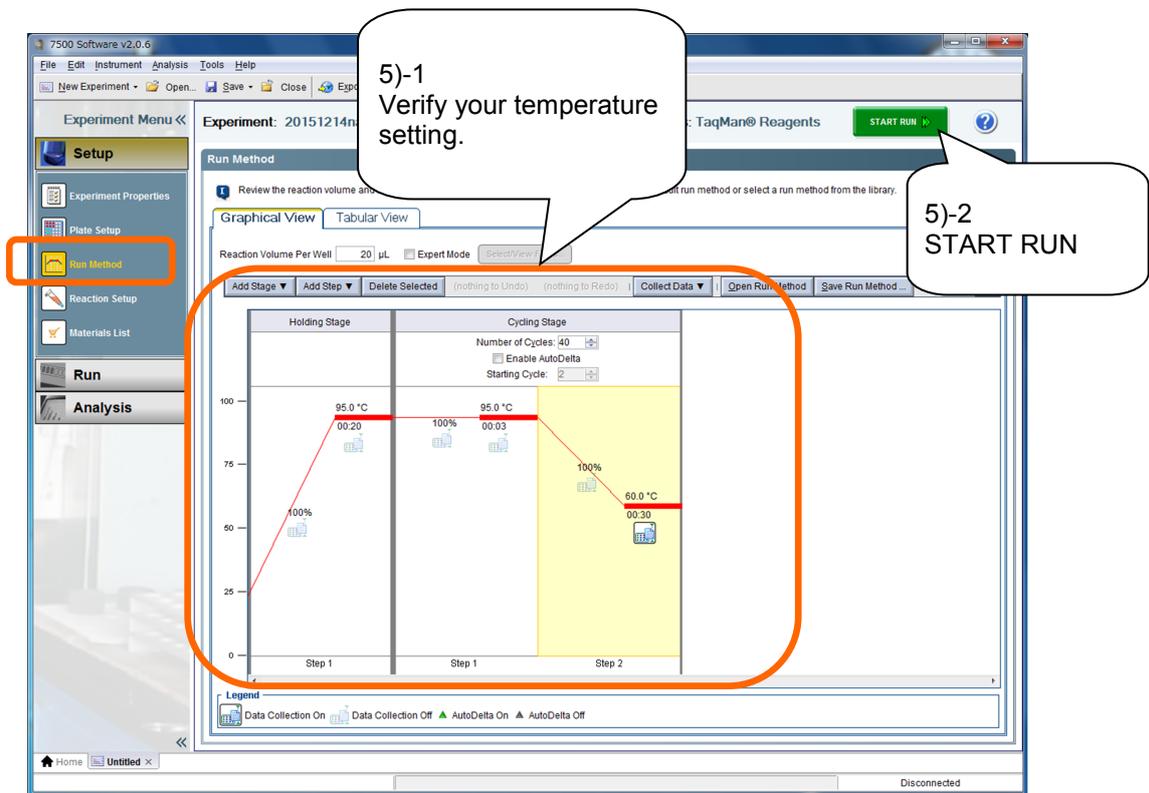
Target genes

Samples

4)-2 Set up your Plate Layout



5) Starting RUN



5)-3 Save the File

The save file window is opened, after clicking on the START RUN button. You should save your file in the following location.

Desktop -> 7500Data(short cut) -> your Lab's folder -> your folder

6) Make sure that the temperature of the heat block begin to rise.

You can see the Temperature Plot on the <RUN> <Temperature Plot> window

